

# ChannelCheck™



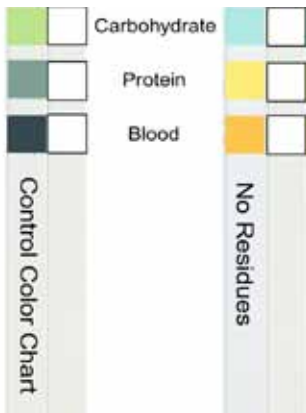
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#### DIRECTIONS:

1. Using at least a 20ml syringe, fill with 10ml of sterile-DI water and 10ml of air.
2. Flush lumen with the sterile-DI water (i.e., sterile water for irrigation) followed by air.
3. Recapture water in a clean (preferably sterile) container (i.e. urine specimen cup).
4. Dip test strip into water being sure that all three pads are completely submerged. Swish the strip in the water for 10 seconds.
4. Remove strip from water and touch side of strip to clean, absorbent surface to wick away excess water.
5. Wait 90 seconds.
6. Compare test strip to the "No Residues" color chart above to interpret results.
7. Should the color on any pad deviate from the "No Residues" this indicates a dirty instrument and it should be re-cleaned and re-tested until test results match the "No Residues" pads.

#### Directions for Control:

1. When opening a new bottle for the first time, a control test should be run (only once per bottle).
2. Check expiration date on control packaging to insure still in-date.
3. Add 1ml of sterile-DI water (i.e., sterile water for irrigation) to the control vial.
4. Recap control vial and shake vigorously to completely rehydrate the soil.
5. Immediately test the control with a strip. If more than 5 minutes has elapsed, result on carbohydrate pad may be different than the Control Chart.
6. Dip test strip into the control vial being sure that all three pads are completely submerged. Swish the strip in the control vial for 10 seconds.
7. Remove strip from the control vial and touch side of strip to clean, absorbent surface to wick away excess liquid.
8. Wait 90 seconds.
9. Compare test strip to the "Control Color Chart" above to interpret results. Colors should closely match those of the Control Color Chart.

**IMPORTANT: KEEP CAP ON TIGHT BETWEEN USES.** Be sure to read insert instructions prior to first use. Be sure to closely follow storage instructions to maximize shelf-life.