Summary
The ChannelCheck™ makes it possible to test a variety of lumened instruments for cleanliness. ChannelCheck™ test strips are comprised of 3 individual reagent pads, each sensitive to a different organic soil: carbohydrate, protein and hemoglobin. These are soils identified by scientific studies as frequently remaining in the lumens of surgical instruments after clinical-use. Also included in this package is a vial of control soil. This soil includes carbohydrate, protein and hemoglobin, the same soils the ChannelCheck™ is designed to test for.

Limits of Testing
Use of these strips can aid in evaluating the efficacy of the cleaning process for individual instruments, but ARE NOT a substitute for vigilance in observation of the state of surgical instruments. Each pad has a specific level of detection (listed below), and instruments may be exposed to other organic and inorganic soils not tested for by the pads. Further, this test is sensitive to organic soils, but is not a measure of microbiological contamination or sterility. Several factors influence interpretation of results - variability in color perception, lighting conditions, and the storage conditions of the product.

Sensitivity of Reagent Pads
Carbohydrate ≥ 210 ug/ml
Protein ≥ 120 ug/ml
Hemoglobin ≥ 0.25 ug/ml

Storage Instructions
For best results, always store ChannelCheck™ bottles tightly capped and in a cool, dry place out of direct sunlight. Note the expiration date on the bottle, which is for the test strips and the expiration date on the packaging for the control soil, which is for the soil. Do not use if either date has expired.
The test-strips are best used by 90 days, once opened and the seal on the test-strip bottle has been broken. After 90 days, the pads MAY change color, before use, indicating a false-positive. IF THE PADS CHANGE COLOR THEY SHOULD NOT BE USED. If the color on the pads remains unchanged the test-strips can still be used. It is IMPORTANT that the test-strips be protected from ambient moisture, light, and heat to guard against altered reagent activity and deterioration. The EXPIRATION DATE printed on the box is the storage life of the test-strips, which can be stored unopened until the EXPIRATION DATE. However, please note that once opened, the 90 days of use should still be prior to the EXPIRATION DATE.

Testing an Instrument for Residuals
1. Fill Syringe with Water: Using at least a 20ml syringe, fill with 10ml of sterile-DI (deionized)water and 10ml of air.
2. Flush Sterile-DI Water Through Channel: This is done by flushing the channel(s) of the instrument with 10mL of sterile-DI water (e.g., sterile water for irrigation) followed by the 10ml of air to facilitate flushing.
3. Recapture Water in the Zip-Lock Bag: Recapture the water in a clean container, such as the supplied zip-lock bag.
4. Dip Test Strip into Water and Swish: Dip the test strip into the recaptured water, being sure that all three pads are completely immersed. Swish the test strip for 10 seconds.
5. Dab Side of Test Strip: Then remove it from the water. Dab the side of the test strip on a clean, absorbent surface to wick away excess water.
6. Wait 90 Seconds: The reagents in the test pads require time to interact with the residual soil, so wait a complete 90 seconds before reading the results.
7. Compare to Color Chart: Compare test strip to the “No Residues” color chart to interpret results.
8. Interpret Results: Should the color on any pad deviate from the “No Residues” this indicates a dirty instrument and it should be re-cleaned and re-tested until test results match the “No Residue” pads.
9. Record Results: On a log sheet, record the results of each pad.
Testing With Control Soil

The first step when opening a new bottle of ChannelCheck™ residual soil test strips is to check the performance of the lot with the included vial of control soil. This will insure that the reagent in each of the test pads has remained active after shipment. This is only done once per bottle and only 2 control vials (1 per bottle) are included.

To test, remove the vial of dehydrated test soil from the box. The test vial holds enough lyophilized test soil to create a single milliliter of test soil.

1. Re-hydrate Soil: To test, remove the vial of dehydrated test soil from the box. The test vial holds enough lyophilized test soil to create a single milliliter of test soil.

2. Shake Vigorously: Shake the vial vigorously, being sure you have a tight seal. Screw the cap back on the vial.

3. Retrieve a Single Test Strip:
   Retrieve a single test soil. Add exactly 1ml of sterile deionized water. After 10 seconds, remove the test strip from the pack. Swirl the test strip in the vial for 10 seconds.

4. Dip Test Strip into Vial: Dip the test strip into the vial, making sure to completely immerse all three test pads into the solution.

5. Swish Test Strip: Swish the test strip in the vial for 1 minute.

6. Dab Side of Test Strip on Absorbent Pad: After 10 seconds, remove the test strip and dab the side of the moistened test pad on a clean, dry absorbent pad, to wick off excess water.

7. Wait 5 minutes: The reagents in the test pads require time to interact with the residual soil, so wait a complete 5 minutes before reading the results.

8. Compare Results to Control Color Chart: After 5 minutes, compare the results to the Control Result Color Chart. The colors of each test pad should closely approximate the colors found on the Control Color Chart.

9. Record Results: On a log sheet, record the results of each pad.

REFERENCES


